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NEWS 19 May 19 Simultaneous left and right truncation added to WSCA  
NEWS 20 May 19 RAPRA enhanced with new search field, simultaneous left and right truncation  
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NEWS 23 Jun 20 2003 edition of the FSTA Thesaurus is now available  
NEWS 24 Jun 25 HSDB has been reloaded  
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NEWS 27 Jul 21 Polymer class term count added to REGISTRY  
NEWS 28 Jul 22 INPADOC: Basic index (/BI) enhanced; Simultaneous Left and Right Truncation available  
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NEWS 30 AUG 13 Field Availability (/FA) field enhanced in BEILSTEIN  
NEWS 31 AUG 15 PATDPAFULL: one FREE connect hour, per account, in September 2003  
NEWS 32 AUG 15 PCTGEN: one FREE connect hour, per account, in September 2003  
NEWS 33 AUG 15 RDISCLOSURE: one FREE connect hour, per account, in September 2003  
NEWS 34 AUG 15 TEMA: one FREE connect hour, per account, in September 2003  
NEWS 35 AUG 18 Data available for download as a PDF in RDISCLOSURE  
NEWS 36 AUG 18 Simultaneous left and right truncation added to PASCAL  
NEWS 37 AUG 18 FROSTI and KOSMET enhanced with Simultaneous Left and Right Truncation

NEWS 38 AUG 18 Simultaneous left and right truncation added to ANABSTR

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FILE COVERS 1907 - 26 Aug 2003 VOL 139 ISS 9  
FILE LAST UPDATED: 25 Aug 2003 (20030825/ED)

This file contains CAS Registry Numbers for easy and accurate substance identification.

=> pronase and virus (w) isolation  
7654 PRONASE  
13 PRONASES  
7661 PRONASE  
(PRONASE OR PRONASES)  
286631 VIRUS  
61896 VIRUSES  
296997 VIRUS  
(VIRUS OR VIRUSES)  
219875 ISOLATION  
947 ISOLATIONS  
220482 ISOLATION

(ISOLATION OR ISOLATIONS)

1021 VIRUS (W) ISOLATION

L1 2 PRONASE AND VIRUS (W) ISOLATION

=> (streptomyces griseus trypsin)

32009 STREPTOMYCES

4221 GRISEUS

64422 TRYPSIN

473 TRYPSINS

64464 TRYPSIN

(TRYPSIN OR TRYPSINS)

L2 61 (STREPTOMYCES GRISEUS TRYPSIN)

(STREPTOMYCES (W) GRISEUS (W) TRYPSIN)

=> virus (w) putification

286631 VIRUS

61896 VIRUSES

296997 VIRUS

(VIRUS OR VIRUSES)

L3 0 PUTIFICATION

0 VIRUS (W) PUTIFICATION

=> Virus (w) purification

286631 VIRUS

61896 VIRUSES

296997 VIRUS

(VIRUS OR VIRUSES)

289567 PURIFICATION

874 PURIFICATIONS

290151 PURIFICATION

(PURIFICATION OR PURIFICATIONS)

249592 PURIFN

232 PURIFNS

249695 PURIFN

(PURIFN OR PURIFNS)

419722 PURIFICATION

(PURIFICATION OR PURIFN)

L4 915 VIRUS (W) PURIFICATION

=> L4 and L2

L5 0 L4 AND L2

=> virus (w) isolation

286631 VIRUS

61896 VIRUSES

296997 VIRUS

(VIRUS OR VIRUSES)

219875 ISOLATION

947 ISOLATIONS

220482 ISOLATION

(ISOLATION OR ISOLATIONS)

L6 1021 VIRUS (W) ISOLATION

=> L6 and L2

L7 0 L6 AND L2

=> pronase

7654 PRONASE

13 PRONASES

L8 7661 PRONASE

(PRONASE OR PRONASES)

=> L6 and L8

L9 2 L6 AND L8

=> L8 and L4

L10 8 L8 AND L4

=> L10 and HAV

1026 HAV

19 HAVS

1033 HAV

(HAV OR HAVS)

L11 0 L10 AND HAV

=> DIS L10 1- IBIB ABS

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L10 ANSWER 1 OF 8 CAPLUS COPYRIGHT 2003 ACS on STN

ACCESSION NUMBER: 1997:270740 CAPLUS

DOCUMENT NUMBER: 126:248758

TITLE: Purification and crystallization of the attachment proteins of enveloped animal viruses using a virosome intermediate

INVENTOR(S): Portner, Allen; Takimoto, Toru

PATENT ASSIGNEE(S): St. Jude Children's Research Hospital, USA; Portner, Allen; Takimoto, Toru

SOURCE: PCT Int. Appl., 40 pp.

CODEN: PIXXD2

DOCUMENT TYPE: Patent

LANGUAGE: English

FAMILY ACC. NUM. COUNT: 1

PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
WO 9709345	A1	19970313	WO 1996-US14187	19960906
W: AL, AM, AT, AU, AZ, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, US, UZ, VN, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM				
RW: KE, LS, MW, SD, SZ, UG, AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI				
AU 9671545	A1	19970327	AU 1996-71545	19960906
PRIORITY APPLN. INFO.:			US 1995-3447P	P 19950908
			WO 1996-US14187	W 19960906

AB A method of purifying the attachment proteins of enveloped viruses in a biol. active form suitable for crystn. and X-ray crystallog. anal. is described. The proteins are incorporated into virosomes by solubilization of the virus with detergent followed by sedimentation of the nucleocapsid and matrix proteins. The supernatant contg. the solubilized attachment proteins and envelope lipids is then treated to remove the detergent with reconstitution of virosomes. The sol. domains of the protein can then be removed by proteolytic cleavage and the residual virosomes removed by sedimentation. The hemagglutinin-neuraminidase (HN) of the Kansas strain of Newcastle's disease virus was purified from allantoic fluid by resuspending the virus at 20 mg/mL in PBS contg. Triton X-100 2 vol.% and incubated at room temp. for 1 h. Nucleocapsid and matrix proteins were pelleted by centrifugation and detergent removed from the supernatant using Bio-Beads. The extracellular domain of the HN was solubilized by treatment with pronase to give a single band on gel electrophoresis.

L10 ANSWER 2 OF 8 CAPLUS COPYRIGHT 2003 ACS on STN

ACCESSION NUMBER: 1991:467248 CAPLUS

DOCUMENT NUMBER: 115:67248  
TITLE: A new method for the purification of the influenza A virus neuraminidase  
AUTHOR(S): McKimm-Breschkin, J. L.; Caldwell, J. B.; Guthrie, R. E.; Kortt, A. A.  
CORPORATE SOURCE: Div. Biomol. Eng., CSIRO, Parkville, 3052, Australia  
SOURCE: Journal of Virological Methods (1991), 32(1), 121-4  
CODEN: JVMEDH; ISSN: 0166-0934  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB A rapid new method for the purifn. of neuraminidase (NA) heads from influenza A virus is described. Virus was pelleted directly from allantoic fluid and was digested with **Pronase**. The cores were removed by centrifugation, redigested and the released NA heads were pooled and concd. The NA was sepd. from all contaminating proteins in a single step on a Superose 12 column. The purified material was suitable for both crystallog. and for the prodn. of monospecific antisera.

L10 ANSWER 3 OF 8 CAPLUS COPYRIGHT 2003 ACS on STN  
ACCESSION NUMBER: 1984:402572 CAPLUS  
DOCUMENT NUMBER: 101:2572  
TITLE: Partial characterization of a transformation-specific glycopeptide in SSV-NP cells  
AUTHOR(S): Thiel, Heinz Juergen; Hafenrichter, Rudolf; Greger, Bernd  
CORPORATE SOURCE: Fed. Res. Cent. Virus Dis. Anim., Tuebingen, D-7400, Fed. Rep. Ger.  
SOURCE: Virology (1984), 134(1), 138-47  
CODEN: VIRLAX; ISSN: 0042-6822  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB An autologous antiserum against simian sarcoma virus-infected nonproducer cells (SSV-NP cells) recognized a SSV transformation-specific glycopeptide (SSV-TrSgp) (Thiel, H. J., et al., 1981). Gel filtration of this component on a Sephadryl S-200 column indicated an apparent mol. wt. at .apprx.200,000. This antigen represented a proteoglycan-like mol., as evidenced by the size of glycopeptides after **Pronase** treatment and by incubation with chondroitinases. The antigenicity of the SSV-TrSgp was completely destroyed after exposure to different proteases. On the other hand, incubation with neuraminidase or chondroitinases degraded the mol. to some extent, but did not affect its antigenicity as measured by immunoprt. Trypsin and EDTA treatment of intact pulse-labeled cells, as well as surface iodination, indicated that the SSV-TrSgp represents a cell membrane-assoccd. mol.

L10 ANSWER 4 OF 8 CAPLUS COPYRIGHT 2003 ACS on STN  
ACCESSION NUMBER: { 1980:123603 CAPLUS  
DOCUMENT NUMBER: 92:123603  
TITLE: Isolation and preliminary characterization of herpes Channel Catfish virus DNA  
AUTHOR(S): Robin, Jean; Rodrigue, Alice  
CORPORATE SOURCE: Fac. Sci., Univ. Sherbrooke, Sherbrooke, QC, J1K 2R1, Can.  
SOURCE: Canadian Journal of Microbiology (1980), 26(2), 130-4  
CODEN: CJMIAZ; ISSN: 0008-4166  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB The DNA of Channel Catfish virus (CCV) was selectively extd. from infected cells with a 5% soln. of Na deoxycholate, deproteinized with Na sarcosinate and **Pronase**, and purified by PhOH extn. followed by equil. d. gradient centrifugation in a CsCl soln. CCV DNA displayed a buoyant d. of 1.715 g/cm<sup>3</sup> in such a soln., as would be expected from a duplex DNA contg. 56.1% guanine plus cytosine. As estd. from both its sedimentation coeff. and length in the electron microscope, CCV DNA is a

linear duplex mol. of .apprx.85 .times. 106 daltons.

L10 ANSWER 5 OF 8 CAPLUS COPYRIGHT 2003 ACS on STN  
ACCESSION NUMBER: 1978:147999 CAPLUS  
DOCUMENT NUMBER: 88:147999  
TITLE: Isolation and study of the electrophoretic mobility of pig influenza virus neuraminidase  
AUTHOR(S): Tolmacheva, V. P.; Daulbaeva, K. D.; Isaeva, E. S.; Chuvakova, Z. K.; Amantaev, S. Zh.  
CORPORATE SOURCE: USSR  
SOURCE: Izvestiya Akademii Nauk Kazakhskoi SSR, Seriya Biologicheskaya (1978), 16(1), 51-5  
CODEN: IKABAR; ISSN: 0002-3183  
DOCUMENT TYPE: Journal  
LANGUAGE: Russian  
AB Neuraminidase from pig influenza virus was isolated by treatment of virus with ether, Tween 20, and pronase followed by centrifugation at 45,000 rpm for 2 h. The products of virus disintegration were pptd. by treatment with formalinized erythrocytes and neuraminidase was found in the supernatant liq. The decrease in yield after erythrocyte treatment indicates that a significant portion of the enzyme is found in complexes with hemagglutinins, which are easily adsorbed on erythrocytes. Polyacrylamide gel electrophoresis showed 2 components with neuraminidase activity; the 1st component decreased significantly after treatment with erythrocytes. The mol. wts. were 250,000 and 200,000 daltons, characteristic for a tetrameric structure of neuraminidase.

L10 ANSWER 6 OF 8 CAPLUS COPYRIGHT 2003 ACS on STN  
ACCESSION NUMBER: 1977:417649 CAPLUS  
DOCUMENT NUMBER: 87:17649  
TITLE: Partial purification and characterization of the potato virus Y helper component  
AUTHOR(S): Govier, D. A.; Kassanis, B.; Pirone, T. P.  
CORPORATE SOURCE: Rothamsted Exp. Stn., Harpenden/Herts., UK  
SOURCE: Virology (1977), 78(1), 306-14  
CODEN: VIRLAX; ISSN: 0042-6822  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB Mg<sup>2+</sup> stabilized potato virus Y helper component during partial purifn. In solns. contg. 0.02M Mg<sup>2+</sup>, the helper component retained most of its activity for 2 days at 4.degree. and for .gtoreq.8 months at -15.degree.. Activity was destroyed on incubation with Pronase or trypsin or by heating for 5 min at 55.degree., but not by incubation with RNase. Incubation with its own antiserum strongly inhibited helper component activity, but antisera to potato virus Y coat protein or inclusion protein had no more effect than a control serum. Filtration through a Sephadex G-200 column resulted in a broad peak of activity which produced many protein-staining bands when electrophoresed on polyacrylamide gel. Gel filtration and ultrafiltration expts. both indicated a mol. wt. of 100,000-200,000. Some helper component activity was retained by aphids allowed to probe into a sucrose soln. for 20 min, showing that the helper component is more firmly bound to the aphid than is the tobacco mosaic virus-poly-L-ornithine complex.

L10 ANSWER 7 OF 8 CAPLUS COPYRIGHT 2003 ACS on STN  
ACCESSION NUMBER: 1977:402099 CAPLUS  
DOCUMENT NUMBER: 87:2099  
TITLE: Highly infectious RNA isolated from cowpea chlorotic mottle virus with low specific infectivity  
AUTHOR(S): Wyatt, S. D.; Kuhn, C. W.  
CORPORATE SOURCE: Dep. Plant Pathol. Plant Genet., Univ. Georgia, Athens, GA, USA  
SOURCE: Journal of General Virology (1977), 35, Pt. 1, 175-80  
CODEN: JGVIAY; ISSN: 0022-1317

DOCUMENT TYPE:

Journal

LANGUAGE:

English

AB Recovery and specific infectivity of infectious RNA from cowpea chlorotic mottle virus of low specific infectivity (14-21 day infections) were greatly improved by using antioxidants during virus purifn. and RNA extn., and by disrupting coat protein with pronase before PhOH-Na dodecyl sulfate extn. Total infectivity of RNA from virus of low infectivity was increased >30-fold. RNA profiles obtained using polyacrylamide gels were then similar for virus with high (4-7 day infections) or low specific infectivity. Low specific infectivity, therefore, seems to be caused by alteration of the coat protein or of the protein-RNA interaction in intact virus particles.

L10 ANSWER 8 OF 8 CAPLUS COPYRIGHT 2003 ACS on STN

ACCESSION NUMBER: 1974:105218 CAPLUS

DOCUMENT NUMBER: 80:105218

TITLE: Preparative isolation of neuraminidase from influenza A viruses (Singapore) 1/57, (Hong Kong) 1/68, and (Leningrad) 99/71

AUTHOR(S): Simanovskaya, V. K.; Vaitkiene, V.; Golubev, D. B.

CORPORATE SOURCE: Vses. Nauchno-Issled. Inst. Grippa, Leningrad, USSR

SOURCE: Voprosy Virusologii (1973), (5), 555-60

CODEN: VVIRAT; ISSN: 0507-4088

DOCUMENT TYPE: Journal

LANGUAGE: Russian

AB The 3 title viruses cultured in the allantoic fluid of chick embryos were used as sources for the prepn. of neuraminidase (I). The extn. method comprised disintegration of the virus with BuOH, ether, and Pronase, pptn. of the S antigen, and gel filtration on Sephadex G-200. The purified I was homogeneous upon polyacrylamide gel electrophoresis. The Singapore strain gave a much higher yield of I, with a much higher sp. activity than either of the other 2 strains. In the process of purification, I acquired a gradually increasing specificity toward the low-mol.-wt. compd. sialyllactose, as opposed to the high-mol.-wt. ovomucin.

=> DIS L9 1- IBIB ABS

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L9 ANSWER 1 OF 2 CAPLUS COPYRIGHT 2003 ACS on STN

ACCESSION NUMBER: 1985:75385 CAPLUS

DOCUMENT NUMBER: 102:75385

TITLE: Pronase treatment of type A/H2N2/ and type B influenza viruses. Isolation of pure neuraminidase heads

AUTHOR(S): Kavaklova, L.; Praskov, D.; Vulkova, B.; Petrunova, S.; Nikolova, Z.; Kotseva, R.

CORPORATE SOURCE: Med. Akad., Sofia, Bulg.

SOURCE: Epidemiologiya, Mikrobiologiya i Infektsiozni Bolesti (1984), 21(4), 23-31

CODEN: EMIBA3; ISSN: 0425-1482

DOCUMENT TYPE: Journal

LANGUAGE: Bulgarian

AB The direct effect of pronase, a protease, was studied on 4 type A and 1 type B influenza virus stains. Pure and active neuraminidase with a very good yield was isolated from strain A/Singapore/1/57/H2N2/. The other 3 type A/H3N2/ strains appeared to have thermolabile and pronase-sensitive neuraminidase and a hemagglutinin relatively resistant to pronase degrdn. The neuraminidase prepns. isolated had low enzyme activity and were hemagglutinin-polluted. Pure neuraminidase with reduced enzyme activity was isolated from strain

B/Singapore 222/79. Monospecific antisera against the pure neuraminidase heads, isolated from A/Singapore/1/57 and B/Singapore 222/79, were obtained. The antisera were used in the double agar diffusion test, aimed at comparing the antigenic identity of type N2 and type B neuraminidases accordingly.

L9 ANSWER 2 OF 2 CAPLUS COPYRIGHT 2003 ACS on STN

ACCESSION NUMBER: 1982:612236 CAPLUS

DOCUMENT NUMBER: 97:212236

TITLE: Isolation of native alpha-virus RNA and several of its physicochemical indexes

AUTHOR(S): Uryvaev, L. V.; Klimenko, S. M.; Samokhvalov, E. I.; Iferov, V. P.

CORPORATE SOURCE: Inst. Virusol. im. Ivanovskogo, Moscow, USSR

SOURCE: Deposited Doc. (1981), VINITI 4502-81, 18 pp. Avail.: VINITI

DOCUMENT TYPE: Report

LANGUAGE: Russian

AB ~~Genomic~~ RNA was isolated from Venezuelan equine encephalomyelitis virus, propagated in chick embryo fibroblasts, by extn. with phenol, treatment with detergent (SDS), or ~~pronase~~ treatment. The yield of RNA was 80-95%. Anal. of RNA in sucrose d. gradient and gel electrophoresis revealed the presence of 3 RNA species with sedimentation coeffs. of 42 S, 28 S, and 18 S. The viral RNA had a mol. wt. of 4.0-4.1 megadaltons and was composed of 11,500-12,500 nucleotides. Electron micrographs of the viral genome are given.